

CAMBRIDGE INTERNATIONAL EXAMINATIONS

Cambridge International Advanced Subsidiary and Advanced Level

MARK SCHEME for the May/June 2015 series

9700 BIOLOGY

9700/52

Paper 5 (Planning, Analysis and Evaluation),
maximum raw mark 30

This mark scheme is published as an aid to teachers and candidates, to indicate the requirements of the examination. It shows the basis on which Examiners were instructed to award marks. It does not indicate the details of the discussions that took place at an Examiners' meeting before marking began, which would have considered the acceptability of alternative answers.

Mark schemes should be read in conjunction with the question paper and the Principal Examiner Report for Teachers.

Cambridge will not enter into discussions about these mark schemes.

Cambridge is publishing the mark schemes for the May/June 2015 series for most Cambridge IGCSE®, Cambridge International A and AS Level components and some Cambridge O Level components.

® IGCSE is the registered trademark of Cambridge International Examinations.

Page 2	Mark Scheme	Syllabus	Paper
	Cambridge International AS/A Level – May/June 2015	9700	52

Mark scheme abbreviations:

;	separates marking points
/	alternatives answers for the same point
R	reject
A	accept (for answers correctly cued by the question, or extra guidance)
AW	alternative wording (where responses vary more than usual)
<u>underline</u>	actual word given must be used by candidate (grammatical variants accepted)
max	indicates the maximum number of marks that can be given
ora	or reverse argument
ecf	error carried forward
I	ignore
mp	marking point (with relevant number)

Page 3	Mark Scheme Cambridge International AS/A Level – May/June 2015	Syllabus 9700	Paper 52
--------	---	------------------	-------------

Question	Expected answer	Extra guidance	Mark
1 (a)	<p><i>independent:</i> <u>concentration</u> of amylase/enzyme ;</p> <p><i>dependent:</i> diameter / area of brown zone ;</p>	<p>I amount R (amylase) extract</p> <p>A radius of brown zone</p> <p>A starch free/digested starch/clear zone</p> <p>A area of brown zone minus well</p> <p>R blue zone</p>	[2]
(b)	<p>any 8 from:</p> <p><i>independent variable</i></p> <p>1 ref. to method of diluting the 0.5 g dm^{-3} amylase/stock amylase solution and to give a minimum of 5 dilutions ;</p> <p>2 ref. to concentrations from 0.5 g dm^{-3} downwards with correct units ;</p> <p>3 use of a <u>control</u> with example ;</p> <p><i>dependent variable</i></p> <p>4 ref. to a suitable method of measuring diameters/width/ radius/area of brown zones ;</p>	<p>1 0.0 and 0.5 g dm^{-3} can be included in the number of dilutions</p> <p>A serial/series/simple/proportional/dilution as method</p> <p>OR a description. Use the formula $C_1V_1 = C_2V_2$ to make...</p> <p>A If the fungal extract is diluted instead of amylase but R mp5</p> <p>2 minimum of 3 other stated values between 0.5 g dm^{-3} and 0.0 g dm^{-3} must correspond to dilution method chosen</p> <p>ecf if no method given</p> <p>A 10 fold or a 50% reduction serial dilution</p> <p>3 A water/0.0 g dm^{-3}/boiled or denatured extract/enzyme</p> <p>4 e.g. using (suitable) ruler/callipers/string and ruler</p> <p>R metre ruler</p> <p>I graticules</p> <p>OR use a (transparent) grid/graph paper and count the number of squares</p> <p>OR take a photograph and measure using one the methods for finding area</p> <p>NOTE: method of measuring must match what they have stated they are measuring.</p>	

<p>5 ref. to testing the (fungal) extract/fungal amylase/fungal enzyme ;</p> <p>6 plot a calibration curve of known concentrations and use it to determine extract concentration ;</p> <p><i>standardising variables (max. 3)</i></p> <p>7 ref. to suitable stated volume/same volume of amylase (in each well) ;</p> <p>8 leave (all plates) for same period of time ;</p> <p>9 method of maintaining at same/constant/optimum/stated temperature ;</p> <p>10 use a buffer to keep the pH of the agar same ;</p> <p>11 same <u>concentration</u> of starch (in the agar plates) ;</p> <p>12 same depth/volume of agar in Petri dish ;</p> <p>13 cover to prevent contamination / evaporation ;</p> <p><i>safety</i></p> <p>14 ref. to low risk investigation/hazard <u>and</u> suitable safety precaution ;</p> <p><i>reliability</i></p> <p>15 ref. to a minimum of three replicates <u>and</u> calculate a mean or identify/eliminate/remove anomalies ;</p>	<p>5 R if dilute the fungal extract</p> <p>6 A compare extract result with results of known concentrations to identify extract concentration OR find approx. range of concentration and then do more at smaller intervals to identify extract concentration</p> <p>7 If a volume is stated – max 1 cm³ I amount/known</p> <p>8 if time stated, minimum 30 min/max 24 hours</p> <p>9 e.g. <u>incubator</u>/constant temp. room A water bath if temp. stated, any single temp. in range 15–65 °C I air conditioning</p> <p>10 e.g. making or adding buffer to agar or starch (solutions) A adding buffer to the amylase solution before using it A if stated, any single pH</p> <p>11 I ref. to other nutrients in agar, I amount</p> <p>12 A depth of agar plate I mass of agar/depth of wells</p> <p>14 fungal/enzyme allergy or fungus/enzyme/iodine/agar is irritant <u>and</u> wearing gloves/eye protection/mask I iodine as an allergen R no risk</p> <p>15 A original and 2 more/several/many/multiple A outliers for anomalies R reduce anomalies</p>	<p>[max 8]</p>
--	---	----------------

Page 5	Mark Scheme	Syllabus	Paper
	Cambridge International AS/A Level – May/June 2015	9700	52

(c)	<p>use a (glucose) biosensor/glucose dipstick or test-strip/named test strip, e.g. clinstix/uristix ;</p> <p>(because) γ-amylase will produce glucose only / β-amylase will produce maltose (mainly) ;</p>	<p>A use Benedict's test and EITHER weigh precipitate OR use colorimeter to find the <u>intensity of blue solution</u> left OR time the <u>first appearance</u> of a colour change</p> <p>A chromatography and stain (to show sugars)</p> <p>A Barfoed's test for monosaccharides</p> <p>I <i>idea that γ-amylase produces more glucose than β-amylase</i> R ora</p>	[2]
(d) (i)	<p><u>1963</u> shown on extract B, plate 5 ;</p> <p><u>1809</u> shown on extract D, plate 3 ;</p>		[2]
(ii)	<p>reject/eliminate/ignore/leave out (affected data from calculations) ;</p>	<p>A repeat until consistent results obtained I repeat unqualified OR repeat to find a mean</p>	[1]

Page 6	Mark Scheme	Syllabus	Paper
	Cambridge International AS/A Level – May/June 2015	9700	52

(iii)	<p>correct values for $\Sigma x = 1820$</p> <p>and $\bar{x} = 303 / 303.3 / 303.3^*$</p> <p>and $(x - \bar{x})^2$;</p> <table border="1" data-bbox="343 433 1230 568"> <tr> <td>$(x - \bar{x})^2$ allowed</td><td>341</td><td>341.3</td><td>341.34</td><td>341.4</td><td>342</td></tr> <tr> <td>$(x - \bar{x})^2$ not allowed</td><td>341.32</td><td>341.2</td><td></td><td></td><td></td></tr> </table> <p>correct value of s from table ;</p> <table border="1" data-bbox="343 671 1096 734"> <tr> <td>correct values of s (\pm)</td><td>8.0</td><td>8.26</td><td>8.27</td><td>8.3</td></tr> </table>	$(x - \bar{x})^2$ allowed	341	341.3	341.34	341.4	342	$(x - \bar{x})^2$ not allowed	341.32	341.2				correct values of s (\pm)	8.0	8.26	8.27	8.3	<p>e.g. <table border="1" data-bbox="1343 274 1736 385"> <tr> <td>Σ</td><td>1820</td><td></td><td>342</td></tr> <tr> <td>\bar{x}</td><td>303</td><td></td><td></td></tr> </table></p> <p>allow ecf for s from wrong value of $\sum (x - \bar{x})^2$</p>	Σ	1820		342	\bar{x}	303			[max 2]
$(x - \bar{x})^2$ allowed	341	341.3	341.34	341.4	342																							
$(x - \bar{x})^2$ not allowed	341.32	341.2																										
correct values of s (\pm)	8.0	8.26	8.27	8.3																								
Σ	1820		342																									
\bar{x}	303																											
(iv)	<p>any 3 from:</p> <p>1 the larger/AW the brown area the more amylase/more enzyme activity ora</p> <p>OR the larger/AW the brown area the more gene copies ;ora</p> <p>2 (person or extract) A had the <u>highest</u>/AW <u>concentration</u> of amylase/(person or extract) F has <u>lowest</u> AW <u>concentration</u> of amylase ;</p> <p>3 (person) A has the highest number of gene copies/(person) F has the lowest number of gene copies ;</p> <p>4 ref. to genetic variation in population (for the production of amylase) ;</p>	<p><i>Ignore ref. to proportionality / positive correlation between enzyme concentration and gene copies</i></p> <p>A strongest/largest/most/ ora weakest/smallest/least</p> <p>A A>B>C>D>E>F</p> <p>I copying results without any conclusion about rank order</p>	[3]																									
		Total: [20]																										

Page 7	Mark Scheme	Syllabus	Paper
	Cambridge International AS/A Level – May/June 2015	9700	52

2 (a)	<p>any 3 from:</p> <p><i>variation in volunteers</i></p> <ol style="list-style-type: none"> 1 body mass/weight ; 2 ref. to use of drugs e.g. medication / self-inflicted ; 3 alcohol consumption ; 4 smoking status ; 5 ref. to ethnicity ; 6 volunteer all have the same 'handedness' ; 7 no medical/named medical condition affecting nerve conduction ; <p><i>variation in method of applying test</i></p> <ol style="list-style-type: none"> 8 same arm tested ; 9 idea of volunteer not moving (during the test) ; 10 electrical charge/coulombs/voltage/potential difference ; 11 same number of volunteers in each age category ; 	<p>Mark the first three given.</p> <p>I environmental factors, e.g. temperature / light / noise in room / time of day</p> <p>I diet</p> <p>I ref. to distance between or position of electrodes / time of charge application</p> <p>7 e.g. Multiple Sclerosis(MS), Myalgic Encephalitis(ME), Muscular Dystrophy(MD), Motor Neurone Disease (MND), spinal bifida, polio</p> <p>A disease of the nervous system</p> <p>9 e.g. at rest / sitting / lying down</p> <p>10 A electrical stimulus</p> <p>I current</p> <p>[max 3]</p>
-------	---	---

(b)	<table border="1" data-bbox="339 207 1230 366"> <tr> <td data-bbox="339 207 444 366" rowspan="2">age/years</td><td data-bbox="444 207 765 366" rowspan="2">mean conduction velocity $\pm S_M$</td><td colspan="2" data-bbox="765 207 1230 255">confidence limits</td></tr> <tr> <td data-bbox="765 255 961 303">lower limit</td><td data-bbox="961 255 1230 303">upper limit</td></tr> <tr> <td data-bbox="339 303 444 366">60–69</td><td data-bbox="444 303 765 366">52.2 ± 0.675</td><td data-bbox="765 303 961 366">50.85</td><td data-bbox="961 303 1230 366">53.55 ;</td></tr> </table>	age/years	mean conduction velocity $\pm S_M$	confidence limits		lower limit	upper limit	60–69	52.2 ± 0.675	50.85	53.55 ;	note: must have both numbers correct for one mark	[1]
age/years	mean conduction velocity $\pm S_M$			confidence limits									
		lower limit	upper limit										
60–69	52.2 ± 0.675	50.85	53.55 ;										
(c) (i)	<p>30–39 and 70–79 OR 40–49 and 70–79 OR 50–59 and 70–79 ; there is no overlap of (confidence) limits / S_M ;</p>	<p>A descriptions, e.g. upper limit of 70–79 does not reach lower limit of 30–39 I error bars overlapping / range(bars) not overlapping I ref. to mean conduction velocity</p>	[2]										
(ii)	<p><i>test:</i> t test ; <i>reason:</i> comparing (two) <u>means</u> / normal distribution / continuous data ;</p>	<p>R continuous variation</p>	[2]										
(iii)	<p>there is no <u>significant</u> difference between the (mean) conduction velocities / NCV of (individuals from) different age groups ;</p>	<p><i>needs to be clear that the significant difference is in the conduction velocity and the not the ages</i> A the difference in (mean) conduction velocities of individuals from different age groups is not <u>significant</u> A stated age categories / women of different ages / young(er) and old(er) women / with age</p>	1										
(d)	<p>large sample size / number of people tested ;</p>	<p>A tested 394 individuals / many people / lots of people ; I sufficient / enough people tested I number of different age categories / ref. to standard error</p>	[1]										
			Total:	[10]									